Photographic key to the Pseudoscorpions of Canada and the adjacent USA

Christopher M. Buddle

McGill University, Department of Natural Resource Sciences, Macdonald Campus, 21,111 Lakeshore Road, Ste-Anne-de-Bellevue, Quebec, H9X 3V9, Canada chris.buddle@mcgill.ca



Abstract. The pseudoscorpion fauna of northern North America is poorly known, in part because of the scarcity of taxonomic resources. This photographic key was developed by combining existing keys with photographs of diagnostic characters from about 80 specimens of pseudoscorpions from North America. The key will classify 35 taxa (to genera and some species) of pseudoscorpions occurring in Canada and the adjacent USA. Some distribution notes are presented, along with information about habitat affinities and general ecology.

Introduction

The order Pseudoscorpiones is a small and curious group of arachnids. They are dorsallyventrally flattened animals, typically less than 5 mm in length and generally light tan to reddish brown to black in colour. Although the order is relatively species-poor (i.e., 3,385 known species, globally (Harvey 2009)), pseudoscorpions are fascinating for their natural history and behaviour (e.g., Weygoldt 1969, Zeh and Zeh 1994, Tizo-Pedroso and Del-Claro 2005), and they exhibit wonderful variability in size and form (Figure 1). Typical habitats for pseudoscorpions include leaflitter, under rocks, within compost piles, under bark and within decaying wood, in caves, and in bird nests. Many species are phoretic on insects or birds (e.g., Haack and Wilkinson 1987), and species such as Chelifer cancroides (L.) are cosmopolitan, typically found in buildings, barns or other humanmade structures. Key references for the pseudoscorpions in North America include Muchmore (1990), Hoff (1949), Nelson (1975), and Chamberlin (1931). These publications, especially Hoff (1949) provide detailed information about collecting specimens and preparing them for taxonomic study.

The most recent phylogenetic treatment of the order was proposed by Harvey (1992), and there has been recent activity on the higher-level taxonomy for the group (e.g., Murienne et al. 2008). Harvey's (2009) on-line catalogue is a key resource for nomenclature and classification, and for its extensive bibliography. However, resources for identification of pseudoscorpions in northern benchmarked the taxonomic status of

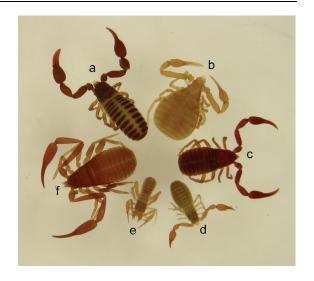


Fig. 1 Selection of pseudoscorpions to illustrate range in body form and size: a. *Dendrochernes* (Chernetidae); b. *Pseudogarypus* (Pseudogarypidae); c. *Wyochernes* (Chernetidae); d. *Microbisium parvulum* (Neobisiidae); e. *Chthonius* (*Ephippiochthonius*) *tetrachelatus* (Chthoniidae); f. *Chelifer cancroides* (Cheliferidae).

North America remain scarce. Buddle (2005) benchmarked the taxonomic status of pseudoscorpions in Canada, and suggests there are 23 valid species known from Canada, with another 20-30 species likely to occur in the country.

Taxonomic keys for pseudoscorpions in Canada are not available; the only key that is workable for North American specimens is that written by Muchmore (1990). This key is an excellent resource, but it is difficult to navigate and

the drawings are difficult for those not trained or experienced in general pseudoscoropion taxonomy. Seeing the characters on actual specimens can help make the taxonomy more accessible; with this in mind, the purpose of this paper is to provide a taxonomic key to genera and some species of pseudoscorpions found in Canada and the adjacent United States of America. This key is not meant to replace other resources, but instead the goal is to provide an additional resource for workers interested in pseudoscorpions. Some information on the general anatomy of pseudoscorpions is included, with the purpose of supplementing information available elsewhere as line drawings (e.g., Hoff 1949, Muchmore 1990). I have also included some notes on the ecology, habitat affinities, and distribution for each taxon, and as more information is gathered in the future, I hope these notes can be developed into formal species pages.

Methods

For each taxon, specimens were either collected in the field, were sent to me over the last several years, or were borrowed from Museums (primarily the Canadian National Collection (CNC), Ottawa, and the Florida State Collection of Arthropods (FSCA)). Whenever possible, I used and examined specimens from northern North America for photographs. William Muchmore also provided many Canadian specimens, and I have relied heavily on many of his slide specimens currently housed at the FSCA. The bulk of the specimens used were slide-mounted, although some characters were better displayed with alcohol specimens and/or only wet specimens were available for study. Slide specimens were photographed using a Leica Infinity1 Camera housed on a Leica DM 2500 compound microscope. For most specimens, 4-6 photographs at different depths of field were taken and subsequently stacked using Combine Zm software (available at http://hadleyweb.pwp.blueyonder. co.uk/CZM/News.htm. Wet specimens were photographed with either the Leica Infinity 1 Camera, or a Nikon Coolpix 4500 Camera, attached to a Nikon SMZ 1500 dissecting microscope. Images were then edited using Adobe Photoshop Elements (Version 4.0).

Over 150 specimens were examined, including all taxa currently represented in Canada. About 80 of these were photographed for generalized anatomy and for diagnostic images for the taxonomic key. For some rarely encountered taxa known only from the adjacent USA, I was

unable to obtain specimens to photograph and these couplets are not accompanied by images (e.g., Aspurochelifer littlefieldi). The key was written by relying heavily on the works of Muchmore (1990), Hoff (1949), and the unpublished manuscripts by M.J. Sharkey. Distributions and ecological information is taken from Muchmore (1990), Harvey (1990, 2009), unpublished reports by M.J. Skarkey, and my personal collections and experience. Unfortunately, some taxa remain unresolved and I have taken a conservative approach when necessary. I have adopted nomenclature proposed by Harvey (2009), unless indicated otherwise, and the key is for a total of 35 taxa (Table 1).

General Anatomy and Taxonomic Characters

The pseudoscorpion body is divided into an anterior cephalothorax (or prosoma) (dorsally, the carapace) and a posterior abdomen (or opisthoma), broadly connected to each other (Figure 2). Males can be differentiated from females, but sexual determination can be difficult. The genital openings occur between the second and third opisthosomal sternites. In general the genital area of females is lightly sclerotized and appears as a bright zone behind the coxae of the legs (Figure 3). In males, this region is relatively dark and more highly sclerotized (Figure 3). Males are also typically smaller than females. Some species also exhibit sexual differentiation in the palps.

The chelicerae contain important taxonomic characters (Figures 2, 4): they are two-segmented structures used to grasp and macerate prey items, and include a fixed and movable finger (Figure 4). Each of the chelicera bears a variety of setae, the placement of which are important for taxonomic purposes. The rallum is a tight grouping on the medial side (Figure 4); the setae of the lateral side are named as follows: exterior seta (es), basal seta (bs), subbasal seta (sbs), interior seta (is), laminal seta (ls), and galeal seta (gs) on the movable finger. A comblike or membraneous serrula occurs along the side of the movable finger, with the *spinneret* or galea at the end, which carries the openings of the silk glands. The third pair of appendages is the pair of long palps or pedipalps, with the coxae of these serving as accessory mouthparts (Figure 2). Beyond the coxa, each palp is comprised of the trochanter, femur, patella, and chela, with a fixed and movable finger. The chela is used for defense and prey capture. The palpal chela usually has numerous teeth along the inner margins, ending with a sharp tip (venedens), which has an opening for the venom gland. The sensory setae

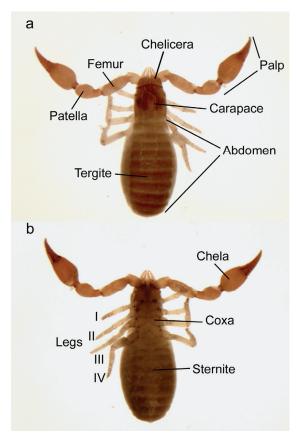


Fig. 2 *Microbisium parvulum* habitus: a. dorsal view; b. ventral view.

(trichobothria) on the movable finger are named as follows (Figure 5), divided by a lateral or exterior series and a medial or interior series: basal (b), subbasal (sb), subterminal (st) and terminal (t). The setae for the fixed finger are named as follows: exterior basal (eb), exterior subbasal (esb), exterior subterminal (est), exterior terminal (et), interior basal (ib), interior subbasal (isb), interior subterminal (ist), and interior terminal (it).

The remaining four pairs of appendages are walking legs (Figure 2), consisting of a coxa, trochanter, basifemur, telofemur, tibia, and one or two tarsal segments. The legs terminate with two claws and an arolium. The abdomen is divided into 11 or 12 segments, with sclerotized *tergites* dorsally and *sternites* ventrally (Figure 2), separated by soft, flexible *intersegmental* and *pleural membranes*. The *spiracles* are found at the lateral edges of the third and fourth sternites.

How to use this key

This dichotomous key is designed for use with both wet specimens and specimens mounted on slides. For most general collections of pseudoscorpions, specimens are typically preserved in 70-95%

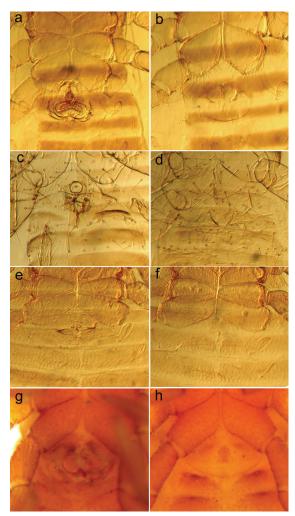


Fig. 3 Representative males (left) and females (right) from different families and species of pseudoscorpions: a,b. Americhernes oblongus (Chernetidae); c,d. Apochthonius moestus (Chthoniidae); e,f. Larca sp. (Larcidae); g,h. Wyochernes sp. (Chernetidae). Photographs from slide specimens except for Wyochernes (specimens in 70% ethanol).

ethanol; typically slide mounts are done only for long-term storage of voucher specimens. When working with wet specimens, it is advised to handle the specimens with small insect pins or fine forceps. For using the key accurately, legs I and IV should be removed (including trochanter but not the coxa), and one chelicera and one chela are typically removed. These are placed separately from the whole specimen on a flat slide with a cover slip propped up with a small piece of fishing line; some structures may need to be cleared for a period of 1-3 days or more (i.e., if heavily sclerotized) with a clearing agent (e.g., lactic acid) if required. It is possible to gently rotate the structure under the

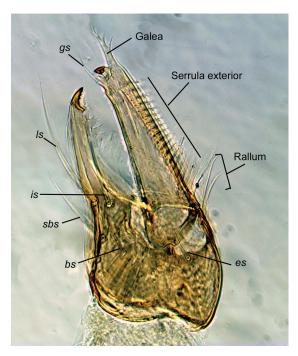


Fig. 4 Acuminochernes sp. chelicera illustrating movable finger with membranous exterior serrula, galea (spinneret), and galeal seta (gs). Fixed finger and base typically contains five setae: exterior seta (es), basal seta (bs), subbasal seta (sbs), interior seta (is) and laminal seta (ls). The rallum is a group of setae near the base of the movable finger.

cover slip by gently moving the cover slip and, provided the fishing line is thin enough, the structure will rotate with ease. In this manner, the correct orientation is achieved. The whole specimen can be viewed by placing in a depression slide with ethanol.

Acknowledgements

Thanks to William Muchmore, GB Edwards (Florida State Collection of Arthropods), Michael Sharkey, Mark Harvey, Terry Wheeler and the Lyman Entomological Museum (McGill University), and the Arachnological Community of Canada for specimens, enthusiasm and moral support. Funding was provided in part from a Discovery Grant (National Science and Engineering Research Council of Canada).

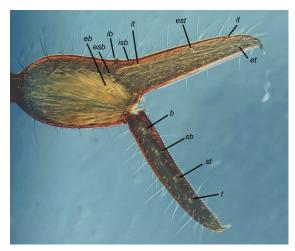
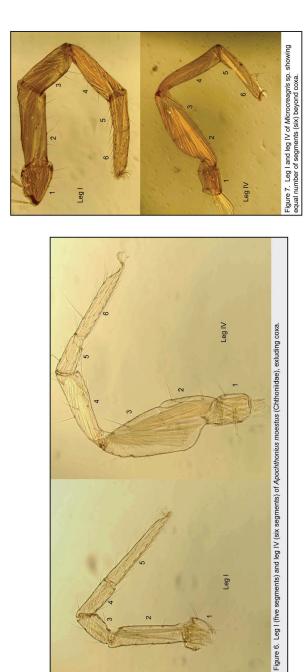


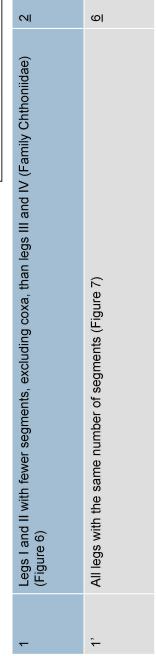
Fig. 5. Palpal chela of *Microcreagris* sp. showing sensory setae (trichobothria). Movable finger contains basal (b), subbasal (sb), subterminal (st) and terminal (t) setae. The fixed finger contains exterior basal (eb), exterior subbasal (esb), exterior subterminal (est), exterior terminal (et), interior subterminal (ist) and interior terminal (it) setae. Trichobothria can be recognized as they are typically longer than other setae, and by the recessed pit at the base of the seta.

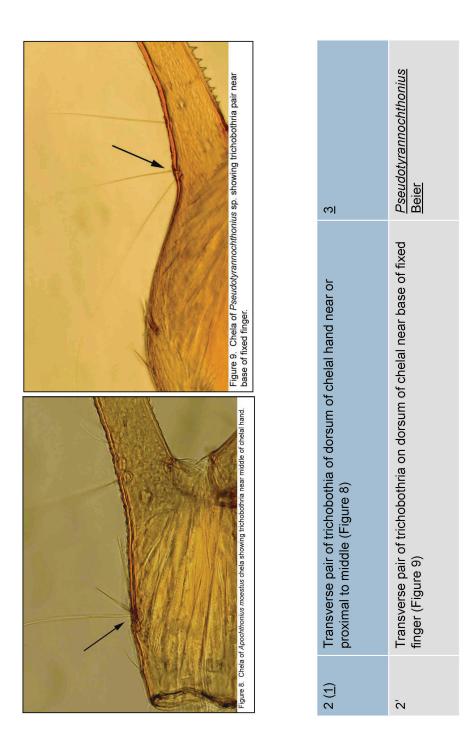
Table 1. Summary of pseudoscorpion taxa that can be determined using the key. Nomenclature follows Harvey (2009).

пагуеў (2009).	
Family	Genus/Species
Chthoniidae	Chthonius (Chthonius) ischnocheles (Hermann)
	Chthonius (Ephippiochthonius) tetrachelatus (Preyssler)
	Apochthonius Chamberlin
	Mundochthonius Chamberlin
	Pseudotyrannochthonius Beier
Syarinidae	Syarinus Chamberlin
Neobisiidae	Americocreagris Ćurčić, Globocreagris Ćurčić, Microcreagris Balzan
	Halobisium occidentale Beier
	Microbisium brunneum (Hagen)
	Microbisium parvulum (Banks)
	Roncus lubricus L. Koch
Larcidae	Larca Chamberlin
Garypinidae	Pseudogarypinus Beier
Pseudogarypidae	Pseudogarypus Ellingsen
Cheiridiidae	Apocheiridium Chamberlin
Chernetidae	Acuminochernes Hoff
	Americhernes Muchmore
	Chernes Menge
	Dendrochernes Beier
	Dinocheirus Chamberlin
	Epactiochernes Muchmore
	Hesperochernes Chamberlin
	Illinichernes Hoff
	Mirochernes dentatus (Banks)
	Parachernes Chamberlin
	Pselaphochernes Beier
	Wyochernes Hoff
Cheliferidae	Aspurochelifer littlefieldi Benedict and Malcolm
	Chelifer cancroides (L.)
	Dactylochelifer Beier
	Haplochelifer philipi (Chamberlin)
	Hysterochelifer Chamberlin
	Idiochelifer nigripalpus (Ewing)
	Parachelifer Chamberlin
	Paisochelifer Hoff

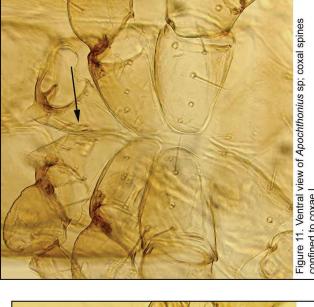
Photographic key to the Pseudoscorpions of Canada and the adjacent USA





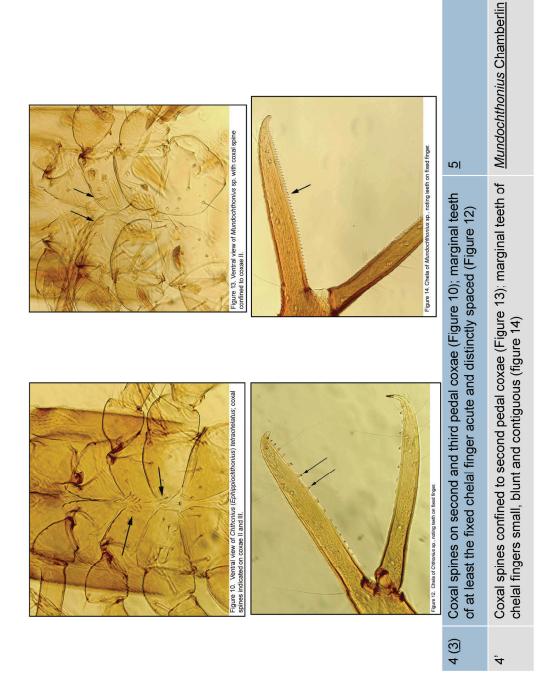


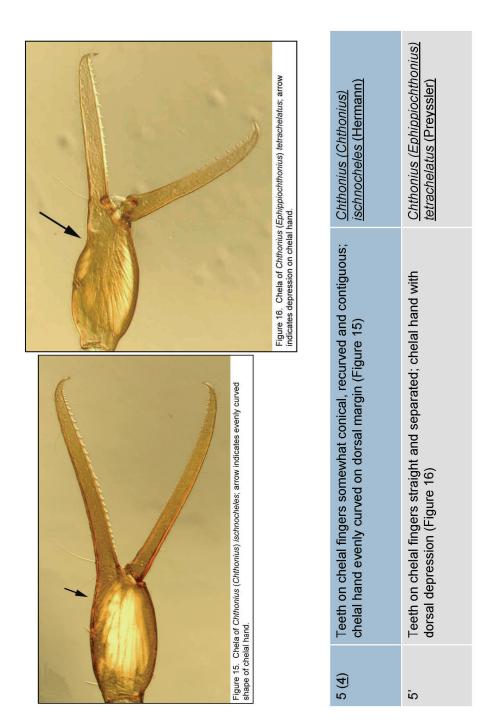


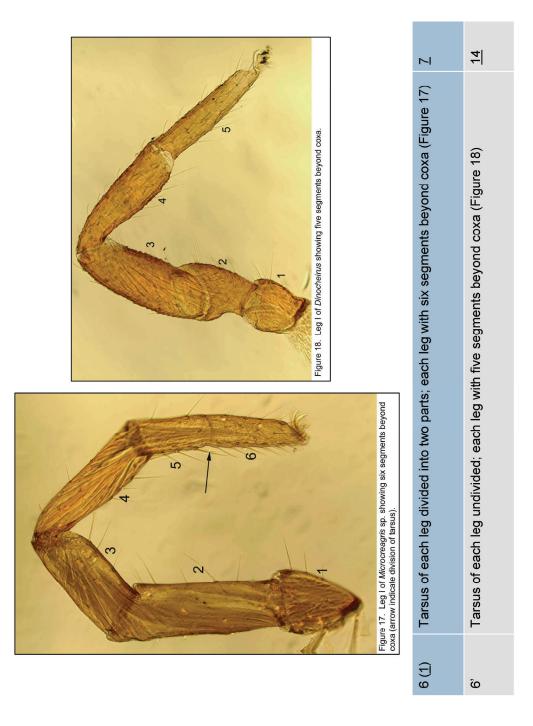




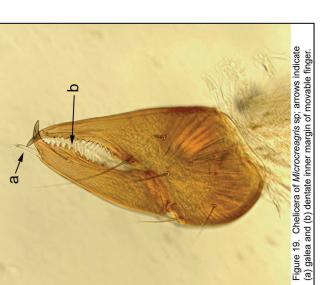
Apochthonius Chamberlin 41 Coxal spines on second and/or third pedal coxae (Figure 10) Coxal spines confined to first coxa (Figure 11) 3 (2) က

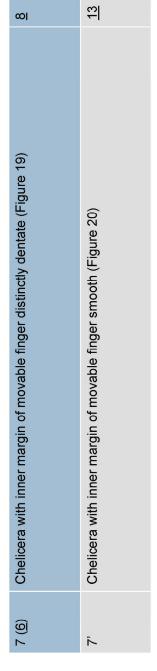


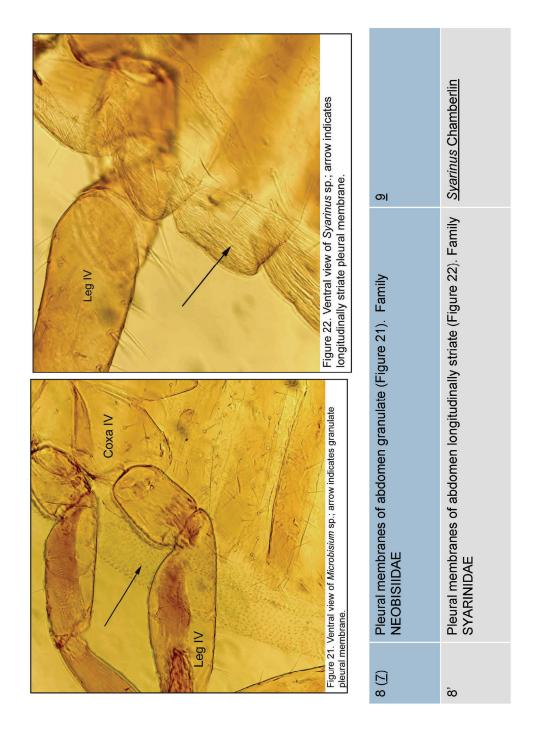


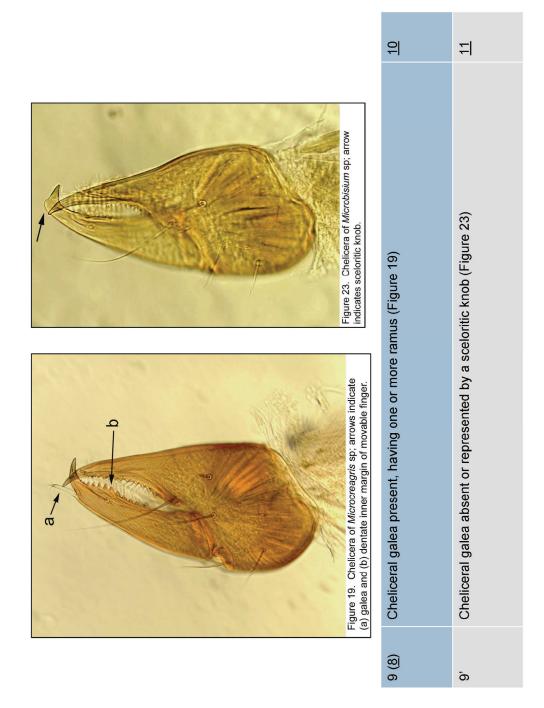


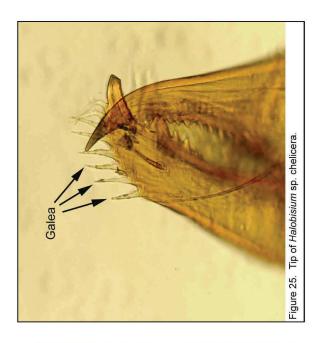


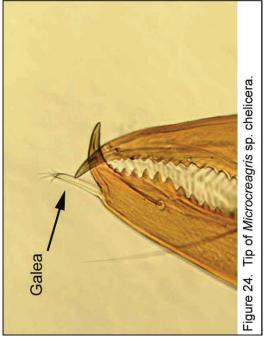




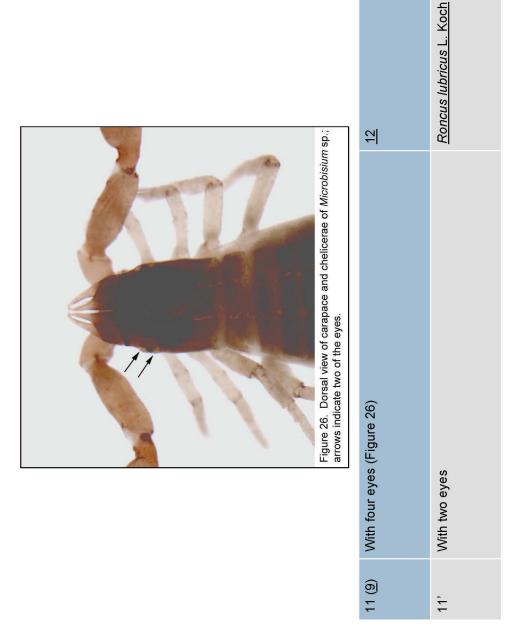








Microcreagris Balzan, Americocreagris Ćurčić and <i>Globocreagris</i> Ćurčić	Halobisium occidentale Beier
Spinneret consisting of a single, usually branched galea (Figure 24)	Spinneret in the form of 8-10 simple galea aligned in a row (Figure 25)
6 0	,o



Microbisium brunneum (Hagen)	Microbisium parvulum (Banks)
12 (11) Chelal length generally greater than 0.70 mm; femur length typically <i>Microbisium brunneum</i> (Hagen greater than 0.40 mm.	Chelal length generally less than 0.70 mm; femur length typically less than 0.40 mm.
12 (11)	12,

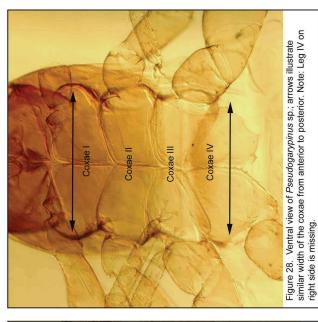
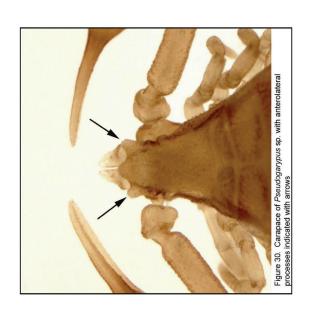


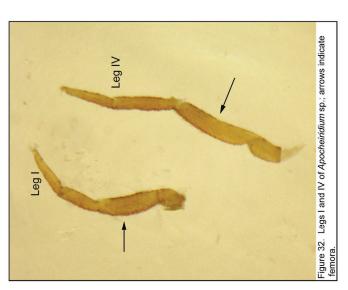
Figure 27. Ventral view of Larca sp.; arrows illustrate the widening of the coxae from anterior to posterior. Coxae IV

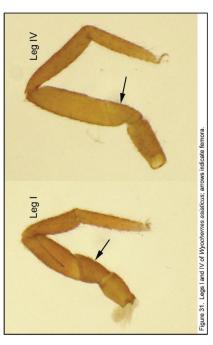
Pseudogarypinus Beier Larca Chamberlin Coxal area with sides approximately parallel (Figure 28). Family GARYPINIDAE Coxal area widened posteriorly (Figure 27). Family LARCIDAE 13 (7) 3,



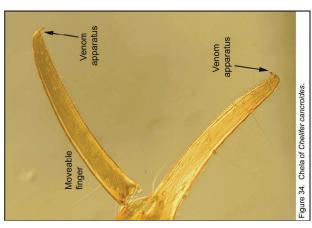


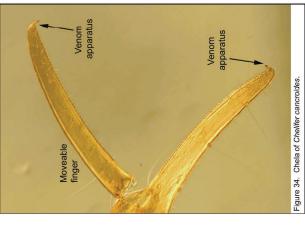
15	Pseudogarypus Ellingsen
With two eyes or without eyes; carapace lacking prominent anterolateral processes (Figure 29)	With four prominent eyes; carapace with distinct anterolateral processes (Figure 30). Family PSEUDOGARYPIDAE
14 (6)	' 4

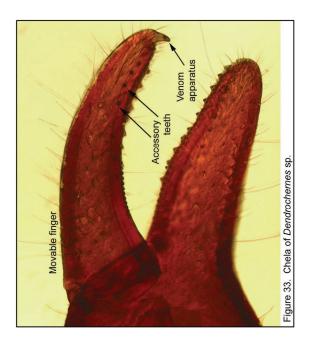




<u>16</u>	Apocheiridium Chamberlin
Femora of legs I and II different in shape than femora of legs III and IV (Figure 31)	Femora of legs I and II similar in shape to femora of legs III and IV (Figure 32). Family CHEIRIDIIDAE
15 (14)	1 5.







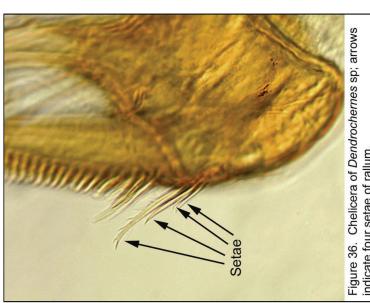
7	 	8
	venom apparatus developed in movable ringer only, may be present as a vestige in fixed finger; accessory teeth usually present on chelal fingers (Figure 33). Family CHERNETIDAE	Venom apparatus equally well developed in fixed and movable fingers; accessory teeth absent from chelal fingers (Figure 34). Family CHELIFERIDAE.
70.7	(<u>CI</u>) 01	16,

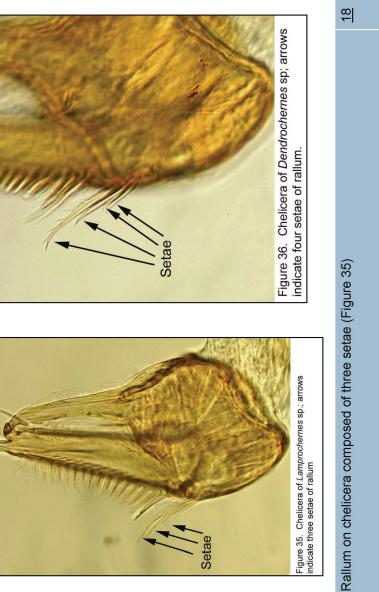
22

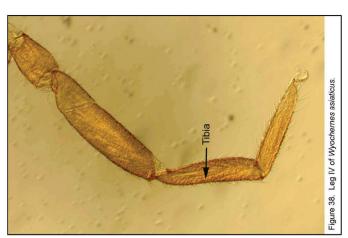
Rallum on chelicera composed of four setae (Figure 36)

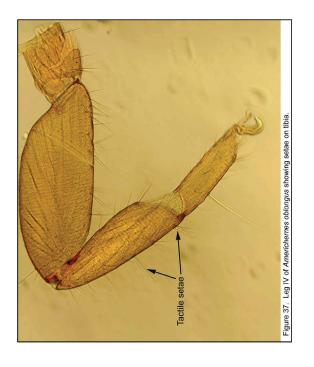
17,

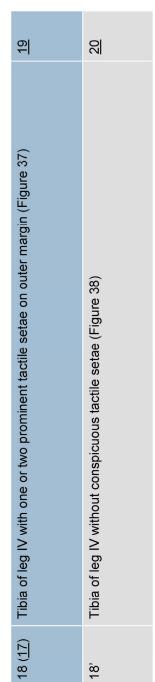
17 (16)

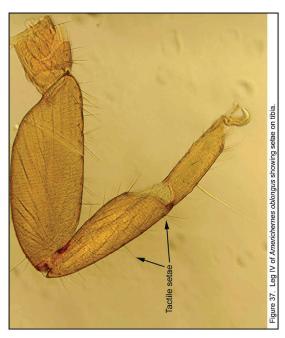


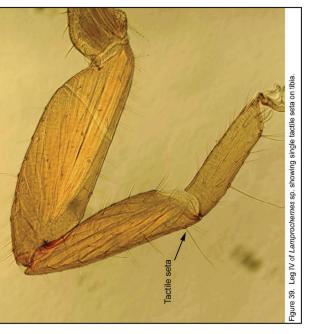




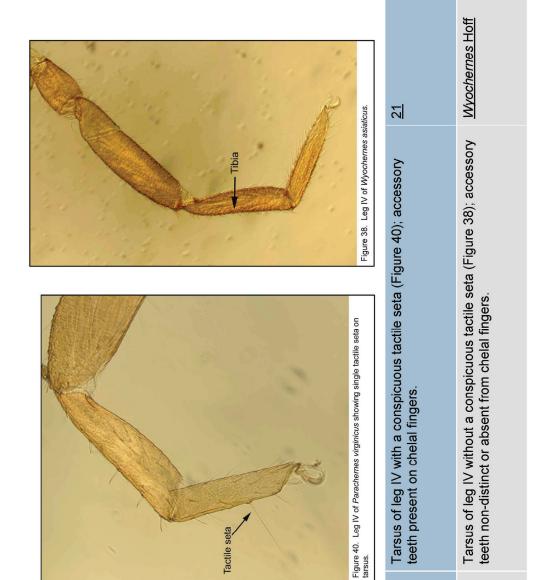






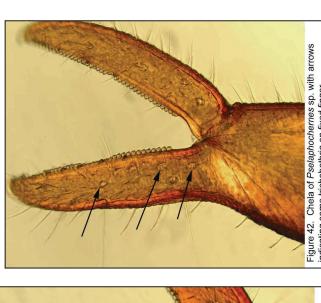


Lamprochernes Tömösvary	<u>Americhernes Muchmore</u>
19 (<u>18</u>) Tibia of leg IV with one tactile seta, near distal end of outer margin (Figure 39)	Tibia of leg IV with two tactile setae, at middle and distal end (Figure 37)
19 (18)	19:



20 (18)

20,



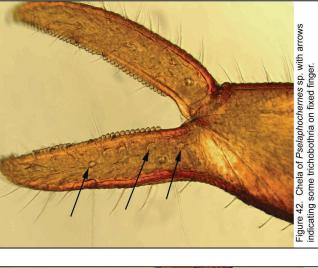


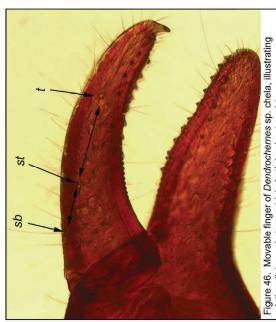
Figure 41. Chela of Parachernes virginicus.

Parachernes Chamberlin	Pselaphochernes Beier
All trichobothria on fixed finger of chela (except et) confined to proximal half of finger (Figure 41)	Trichobothria on fixed finger of chela of standard arrangement (Figure 42)

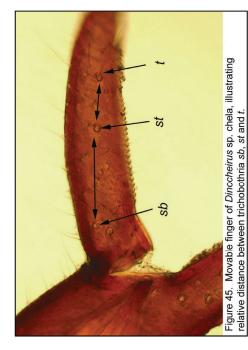
21 (20)

7,









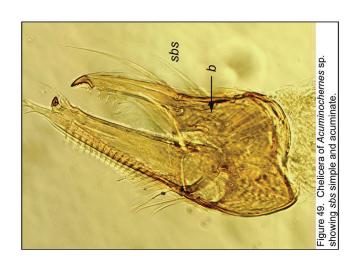
<u>24</u>	Dendrochernes Beier
23 (<u>22</u>) Movable chelal finger with trichobothrium <i>st</i> nearer to <i>t</i> than to <i>sb</i> (Figure 45)	Movable chelal finger with trichobothrium st nearer to sb than to t (Figure 46)
23 (22)	23,

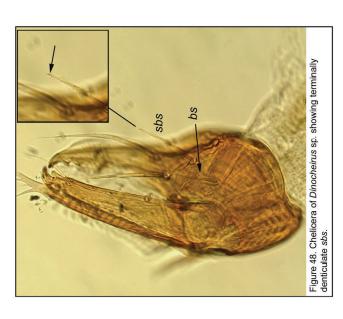






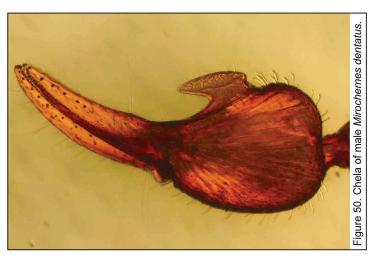
<u>25</u>	Epactiochernes Muchmore
Tactile seta on tarsus of leg IV distinctly distal to middle in position (Figure 47)	Tactile seta on tarsus of leg IV mid-length in position (Figure 43)
24 (<u>23)</u>	24'





<u>76</u>	Acuminochemes Hoff
Hand of chelicera with seta b acuminate and ${\it sb}$ terminally denticulate (Figure 48)	Hand of chelicera with both $\it b$ and $\it sb$ simple and acuminate (Figure 49)
25 (<u>24)</u>	25'





	(Banks)	1) <u>Dinocheirus Chamberlin</u>
I plade to object to be a second beneath the terminate of the second to	Male with profillient, andi-shaped process on medial side of chera hand (Figure 50)	Males without a process on the medial side of chelal hand (Figure 51)
(30) 9	(<u>C</u>)	, 9

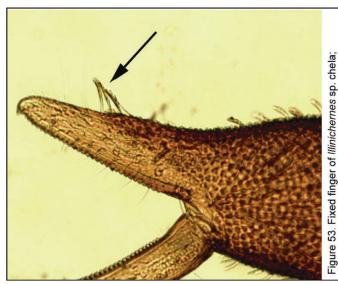
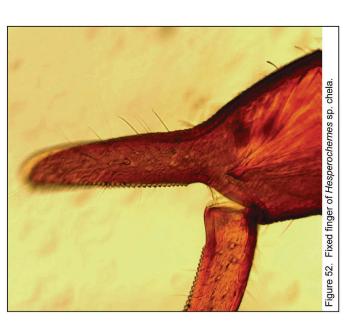
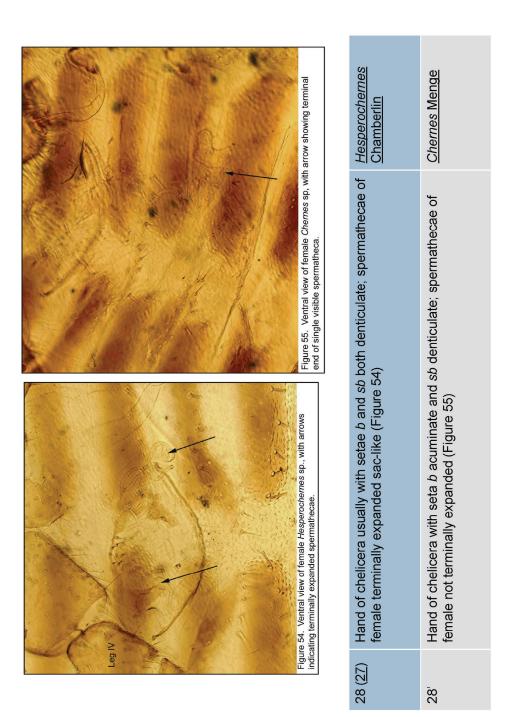
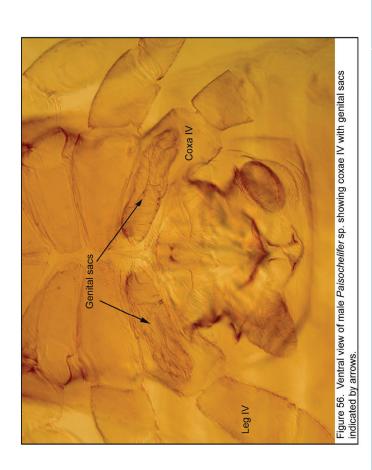


Figure 53. Fixed finger of *Illinichernes* sp. chela; arrow indicates clavate and elongate setae.

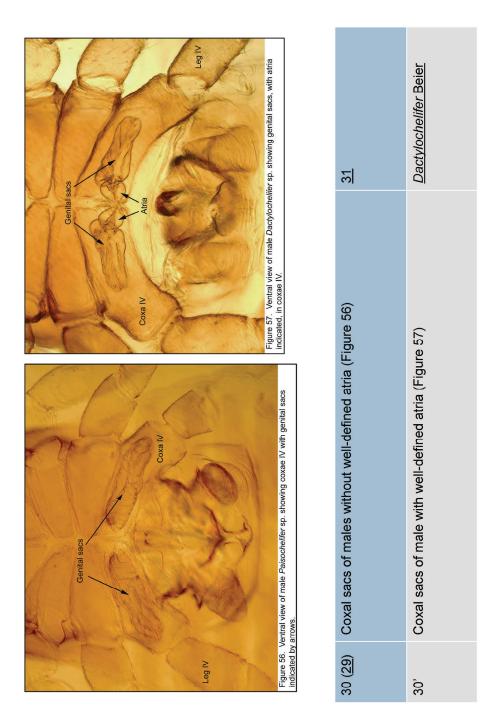


<u>128</u>	Illinichernes Hoff
Setae of palps and tergites regular in form; setae on fixed palpal chelal finger all similar in length and form (Figure 52)	Setae of palps and tergites large, pinnate and leaflike; several long, clavate setae near centre of outer margin of fixed chelal finger (Figure 53)
27 (<u>22</u>)	27'





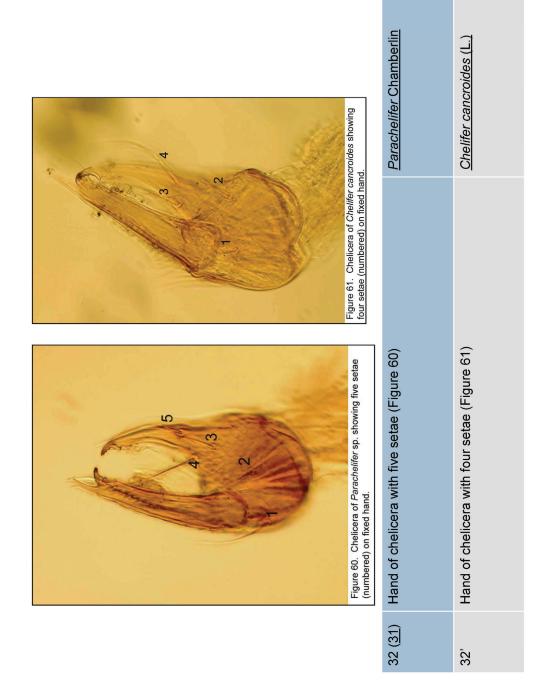
Haplochelifer philipi (Chamberlin) 30 Male with genital sacs in coxae of legs IV (Figure 56); female with median cribriform plates smaller in diameter than diameter of anterior tracheal trunk. Male without genital sacs in coxae of legs IV; female with median cribriform plates as large in diameter as diameter of anterior tracheal trunk. 29 (16) 29,

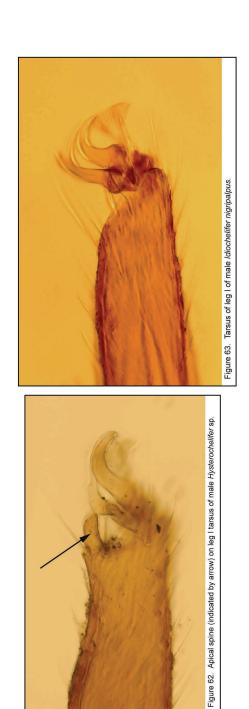


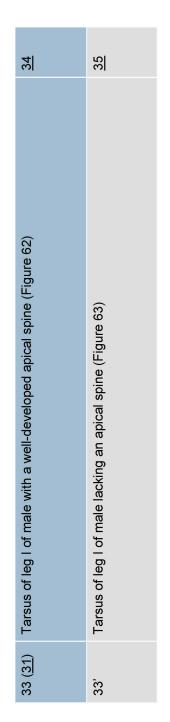


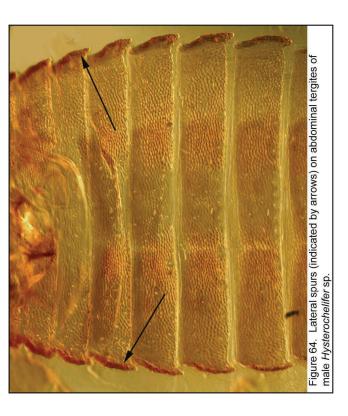


32	83
31 (<u>30)</u> Tarsal claws of leg IV bifid or with an accessory tooth (Figure 58)	Tarsal claws of leg IV simple and without accessory tooth (Figure 59)
31 (30)	. . 23.

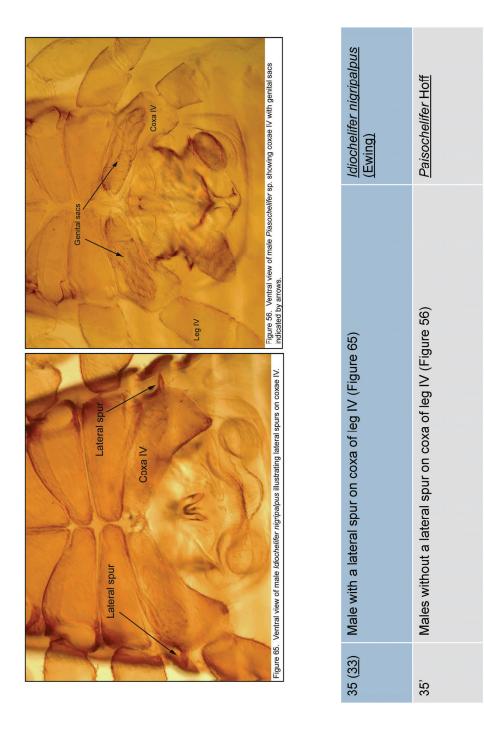








Hysterochelifer Chamberlin	Aspurochelifer littlefieldi Benedict and Malcolm
Male with well developed lateral spurs on anterior abdominal tergites (Figure 64).	Male without lateral spurs on abdominal tergites.
34 (33)	34,



Pseudotyrannochthonius Beiel Chthoniidae

Taxonomy and distribution:

- Four Nearctic species, two likely to occur in western Canada:

P. gracilis Benedict & Malcolm: WA

• P. incognitus (R.O. Shuster): WA, ID

Ecology: leaf litter, soil

Chthoniidae *Apochthonius* Chamberlin

- Taxonomy and distribution:
- Nearctic genus; 24 described species, two Canadian records:
- A. minimus R.O. Shuster: BC
- A. moestus (Banks): ON, widespread species in USA, probably occurs across Canada
- Ecology: moist litter

Chthoniidae *Mundochthonius* Chamberlin

Taxonomy and distribution:

- 10 Nearctic species, two occur or are likely to occur in Canada:

• M. rossi Hoff: MB

• M. pacificus (Banks): western North America

Ecology: moist, cool litter

Chthonius (Chthonius) ischnocheles Chthoniidae (Hermann)

- Taxonomy and distribution:
- Several sub-species recognized
- Holarctic distribution: known from eastern USA and likely occurs in eastern Canada
- Ecology: moist litter, organic debris

Chthoniidae Chthonius (Ephippiochthonius) tetrachelatus (Preyssler)

Taxonomy and distribution:

and eastern USA and known from Canada: Holarctic distribution; widespread in central ON, NS

Ecology: moist litter, organic debris

Syarinus Chamberlin Syarinidae

- Taxonomy and distribution:
- Six recognized species, five of which occur in North America
- Three species known from Canada
- S. enhuycki Muchmore: ON, central and eastern USA
- S. obscurus (Banks): BC, SK, YK and western USA
- S. palmeni Kaisila: NF
- Ecology: under rocks, moist litter, deep in soil

Neobisiidae Americocreagris Ćurčić Globocreagris Ćurčić Microcreagris Balzan

- Taxonomy and distribution:
- (sensu lato) and taxonomic status of the group remains - various genera have been proposed for Microcreagris unstable
- Several species occur in North America and likely occur in Canada
- Americocreagris columbiana (Chamberlin): OR, WA
- Globocreagris theveneti (Simon): AK, AZ, CA, OR, WA
- Microcreagris tacomensis (Ellingsen): WA
- Ecology: cool, moist littter, under rocks and stones

Halobisium occidentale Beie Neobisiidae

- Taxonomy and distribution:
- Only representative of Halobisium in North America
- Known in western USA (AK, CA, OR, WA) and Canada (BC)
- Ecology: littoral species, found under rocks, and in salt marshes

Neobisiidae Roncus Iubricus L. Koch

Taxonomy and distribution:

Widespread in Europe, introduced to North America at least twice: NY, MA

greenhouses; in Europe, tends to occur in Ecology: in North America, known from dry deciduous leaf litter

(Hagen Microbisium brunneum Neobisiidae

- Taxonomy and distribution:
- Nearctic species, widespread and common in North America, including across Canada: AB, MB, NF, NS, QC, ON
- Occurs in northern latitudes
- Difficult to distinguish from M. parvulum
- forests with acidic soils; males are rare boreal forests litter, including bogs and Ecology: in Canada, typically found in

(Banks Microbisium parvulum Neobisiidae

- Taxonomy and distribution:
- across USA and southern Canada: MB, NS Nearctic species, widespread and common ON, QC
- Difficult to distinguish from M. brunneum
- Ecology: deciduous forest litter and soil; males are rare

Larca Chamberli Larcidae

- Taxonomy and distribution:
- Occurs in Europe and North America
- Four species in North America
- L. notha Hoff: Canada (SK), USA (OR, CO)
- L. granulata (Banks) widespread in north-eastern USA and likely occurs in Canada
- Ecology: occurs in litter, frass, mammal

Garypinidae Pseudogarypinus Beier

- Taxonomy and distribution:
- three described species, two occur in southern and western USA
- P. frontalis (Banks) known from WA and ID and likely occurs in western Canada
- Ecology: dry litter, under stones, bird and mammal nests

Pseudogarypidae *Pseudogarypus* Ellingsen

- Taxonomy and distribution:
- Six known species reported from USA and Canada
- P. banksi Jacot known from QC and NS
- Ecology: under rocks, tree holes, rotten wood, stumps

Cheiridiidae *Apocheiridium* Chamberlin

- Taxonomy and distribution:
- Widespread genus, occurs on multiple continents
- At least six species known from USA
- One record known for Canada: Alberta
- Ecology: small body size; varied habitats including potential phoresy; known from bodies of noctuid moths

Tömösvárv Chernetidae Lamprochernes

- Taxonomy and distribution:
- Widespread genus, with three species known from the Nearctic
- L. minor Hoff occurs in central USA and Canada: BC, QC, ON, AB
- Ecology: moist, organic debris; phoretic on insects

Chernetidae *Americhernes* Muchmore

Taxonomy and distribution:

Large genus

Four species known from USA and one occurs in Canada:

· A. oblongus (Say): AB

Ecology: under bark of logs and trees

Wyochernes Hoff Chernetidae

- Taxonomy and distribution:
- Several species and subspecies described from North America and Asia
- Two species occur in North America

W. asiaticus (Redikorzev): YK, AK

- Beringian species
 - W. hustoni Hoff: WY
- Ecology: under stones and rocks at high elevations and latitudes; dry creek beds

Chernetidae *Paracherne*s Chamberlin

- Taxonomy and distribution:
- Diverse and primarily neotropical genus,
- one species, P. virginicus (Banks), occurs in north-eastern USA and probably in adjacent Canada
- Ecology: under bark of trees and logs; leaf

Pselaphochernes Beiel Chernetidae

- Taxonomy and distribution:
- Diverse genus, occurs in North America and Europe
- Three species known to occur North America with two possibly occurring in Canada:
- P. parvus Hoff: central USA
- P. scorpionides (Hermann): north-eastern USA
- Ecology: moist litter, compost

Chernetidae *Dendrocherne*s Beier

- Taxonomy and distribution:
- Holarctic genus, four described species
- One Palearctic species (sensu lato): D. cymeus (L. Koch)
- Three Nearctic species, and one undescribed species from Quebec
- D. crassus Hoff: USA (CO, NM, OR), Canada (BC)
- D. instabilis (Chamberlin): MT
- D. morosus (Banks): USA (FI, NY, MI) and Canada (ON, SK)
- Ecology: under bark, or phoretic on insects (e.g., Cerambycidae beetles)

Epactiochernes Muchmore Chernetidae

- Taxonomy and distribution:
- Two Nearctic species described
- E. tristis (Banks): eastern seaboard (USA), likely occurs in eastern Canada
- Ecology: littoral habitats, under rocks and debris on shorelines

Chernetidae *Acuminochernes* Hoff

- Taxonomy and distribution:
- Two Nearctic species described, both may occur in Canada
- A. crassopalpus (Hoff): widespread in USA
- · A. tacitus Hoff: CO, MI
- Ecology: tree holes, decayed logs, often associated with birds and mammals

Mirochernes dentatus (Banks Chernetidae

- Taxonomy and distribution:
- Monotypic genus found across central USA probably occurs in Canada
- Ecology: decayed trees, often associated with small mammals

Chernetidae *Dinocheiru*s Chamberlin

- Taxonomy and distribution:
- Widespread holarctic genus
- 16 species known from USA, with several species likely occurring in Canada
- D. pallidus (Banks): central and eastern North America
- D. serratus (Moles): western USA
- Undescribed species collected from Alberta
- sometimes associated with animal nests Ecology: leaf-litter, organic debris

Chernetidae Illinichernes Hoff

- Taxonomy and distribution:
- Two Nearctic species
- I. distinctus Hoff: occurs in central and eastern USA and likley occurs in Canada
- Ecology: tree holes, decaying logs

Chernetidae *Hesperochernes* Chamberlin

Taxonomy and distribution:

– Widespread and diverse genus in North America

- Two species known from Canada

H. canadensis Hoff: AB, ON, SK
H. tamiae Beier: QC, ON, AB

Ecology: organic debris, often associated with mammals

Chernes Menge Chernetidae

- Taxonomy and distribution:
- Widespread, diverse, occurs in Europe, Asia, Africa and Americas
- Several widespread species in North America
- C. lymphatus (Hoff): occurs in ON
- litter, under bark and phoretic on insects Ecology: typically found in moist forest

Cheliferidae *Haplochelifer philipi* (Chamberlin)

Taxonomy and distribution:

Monotypic genus, occurs in western USA and likely occurs in western Canada

Ecology: leaf litter

Dactylochelifer Beiel Cheliferidae

- Taxonomy and distribution:
- Widespread and diverse genus, with two species occuring in North America
- D. copiosus Hoff: widespread in USA, Canada (NS, BC)
- D. silvestris Hoff: western USA and Canada (AB,
- Ecology: dry leaf litter, organic debris

Cheliferidae *Parachelifer* Chamberlin

Taxonomy and distribution:

- Widespread in Americas

Eight species known from USA, generally occurring in southern USA except:

P. monroensis Nelson: MI

P. montanus Chamberlin: MT

Ecology: under tree bark

Chelifer cancroides (L.) Cheliferidae

- Taxonomy and distribution:
- Monotypic genus, with cosmopolitan distribution
- Occurs across North America, including most regions of Canada
- association with humans and/or domestic Ecology: almost always found in close animals (homes, barns and other

Cheliferidae *Hysterochelifer* Chamberlin

Taxonomy and distribution:

- Holarctic, and widespread genus, with three species known from USA

 H. fuscipes (Banks) and H. proprius Hoff may occur in central or western Canada Ecology: known from typically dry habitats

Aspurochelifer littlefieldi Benedict and Malcolm Cheliferidae

- Taxonomy and distribution:
- Monotypic genus, known from western USA including WA, OR and may occur in western Canada
- **Ecology: forest litter**

diochelifer nigripalpus Cheliferidae

- Taxonomy and distribution:
- Monotypic genus, distributed across cenrral USA and likely occurs in Canada
- Ecology: typically found under bark

Paisochelifer Hoff Cheliferidae

- Taxonomy and distribution:
- Two described species both occurring in North America
- P. callus (Hoff): central USA and Canada (ON, NS, NB)
- Ecology: moist litter, bird nests

References

- Buddle, C.M. 2005. A primer on pseudoscorpions and taxonomic status in Canada. Newsletter of the Biological Survey of Canada (Terrestrial Arthropods) 24: 12-16.
- Chamberlin J.C. 1931. The arachnid order Chelonethida. Stannford Univ. Publ. Univ.Serv. Biol. Sci. 7: 1-284
- Haack, R.A., and R.C. Wilkinson 1987. Phoresy by *Dendrochernes* Pseudoscorpions on Cerambycidae (Coleoptera) and Aulacidae (Hymenoptera) in Florida. Am. Midl. Nat. 117: 369-373.
- Harvey, M.S. 1990. Catalogue of the Pseudoscorpionida (Mahnert V, ed). Manchester Univ. Press.
- Harvey, M.S. 1992. The phylogeny and classification of the Pseudoscorpionida (Chelicerata: Arachnida). Inverteb. Taxon. 6: 1373-1435.
- Harvey, M.S. (2009). Pseudoscorpions of the World, version 1.2. Western Australian Museum, Perth. http://www.museum.wa.gov.au/arachnids/pseudoscorpions/
- Hoff, C.C. (1949) The pseudoscorpions of Illinois. Bull. Illinois Nat History Survey 24: 413-498.
- Muchmore, W.B. 1973. Ecology of pseudoscorpions a review. Pp 121-127, in Proceedings of the First Soil Microcommunites Conference (Dindall DL, ed.) Syracuse NY.
- Muchmore, W.B. 1990. Pseudoscorpionida. Pp 503-527, in Soil Biology Guide (Dindal DL, ed). John Wiley & Sons.
- Murienne, J., M.S. Harvey, and G. Giribet. 2008. First molecular phylogeny of the major clades of Pseudoscorpiones (Arthropoda: Chelicerata). Molecular phylogenetics and evolution. 49: 170-184.
- Nelson S, Jr. (1975) A systematic study of Michigan Pseudoscorpionida (Arachnida). Am. Midl. Nat. 93: 257-301.
- Tizo-Pedroso, E., and K. Del-Claro. 2005. Matriphage in the neotropical pseudoscorpion *Paratemnoides nidificator* (Balzan 1888) (Atemnidae). Journal of Arachnology 33: 873-877.
- Weygoldt, P. 1969. The biology of Pseudoscorpions. Harvard Univ. Press, Cambridge.
- Zeh, D.W., and J.A. Zeh. 1994. When morphology misleads interpopulation uniformity in sexual selection masks genetic-divergence in harlequin beetle-riding pseudoscorpion populations. Evolution 18: 1168-1182.